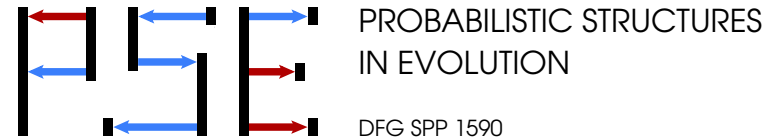


COLLABORATIVE RESEARCH CENTER 1310

## Predictability in Evolution



## Evolutionary pathways to antibiotic resistance

Joachim Krug  
Institute for Biological Physics  
University of Cologne

“Statistical Biological Physics”, ICTS Bangalore, December 9, 2020

# Evolution of antibiotic resistance

- Resistance evolution is a universal response of microbial pathogens to biomedical interventions
- Evolutionary considerations are key for developing strategies that prevent or delay resistance evolution in clinical or environmental settings
- At the same time microbial resistance evolution serves as a model system for addressing broader questions of evolutionary theory

# Evolution of antibiotic resistance

- Resistance evolution is a universal response of microbial pathogens to biomedical interventions
- Evolutionary considerations are key for developing strategies that prevent or delay resistance evolution in clinical or environmental settings
- At the same time microbial resistance evolution serves as a **model system** for addressing broader questions of evolutionary theory

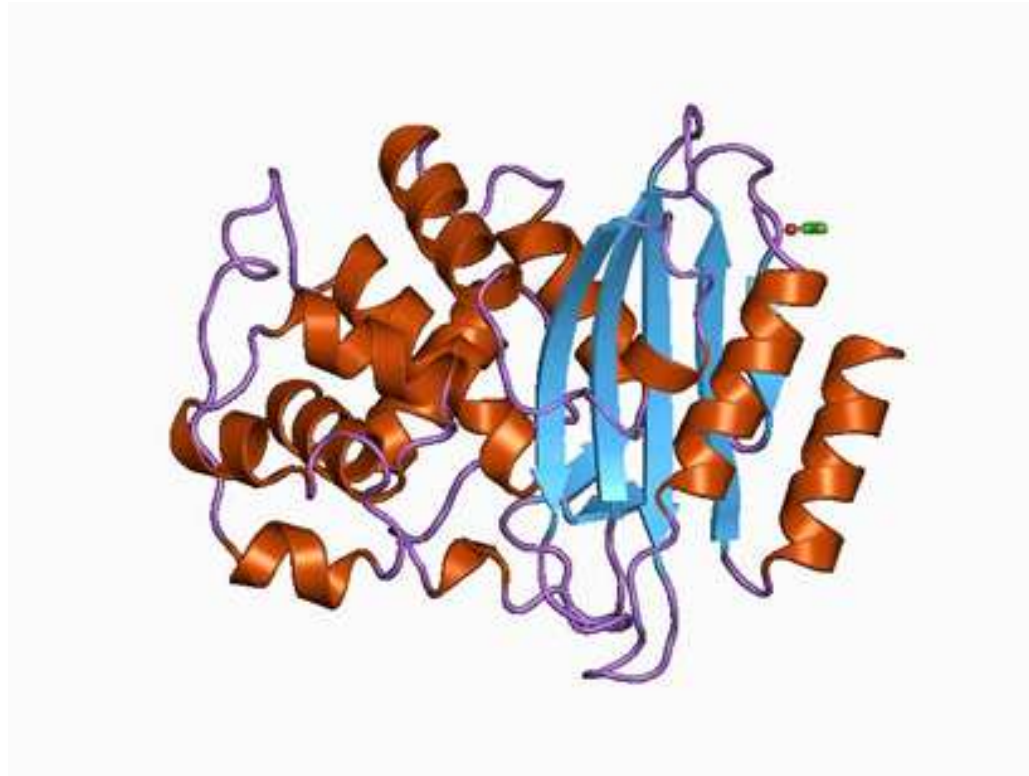
## Two case studies

- The resistance landscape of TEM-1  $\beta$ -lactamase  
joint work with de Visser lab (Wageningen)
- Concentration-dependent evolution of ciprofloxacin resistance  
joint work with Bartek Waclaw and Rosalind Allen (Edinburgh)

# Quantifying antibiotic effects

- The effect of a drug is quantified by the **dose-reponse curve**, the growth rate of a (large) bacterial population as a function of drug concentration
- At the **minimal inhibitory concentration (MIC)** the growth rate drops to zero
- Resistance mutations increase the MIC
- The combination of multiple mutations along an **evolutionary pathway** leads to highly resistant strains

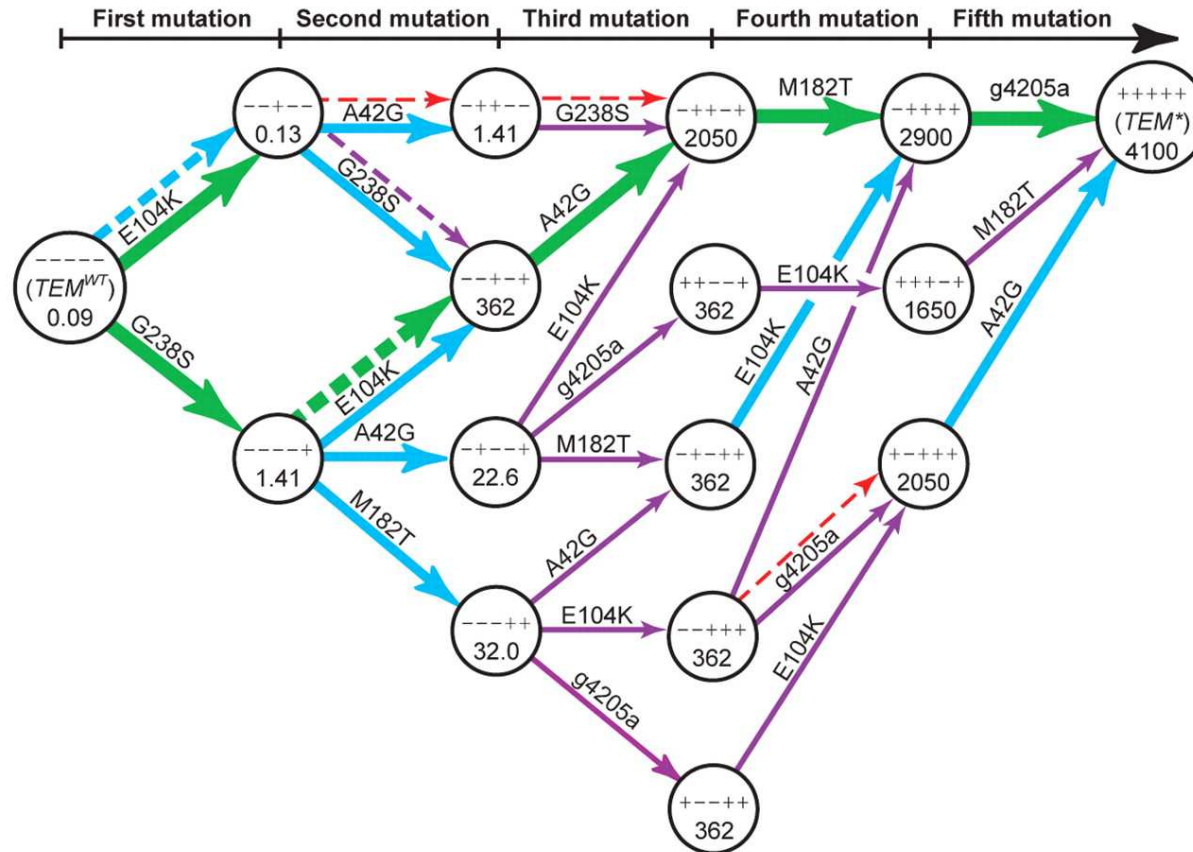
# TEM-1 $\beta$ -lactamase



- $\beta$ -lactam antibiotics such as penicillin target cell wall synthesis
- TEM-1  $\beta$ -lactamase confers resistance against ampicillin to *E. coli*
- Experiments study adaptation to novel antibiotic cefotaxime

# Pathways to TEM-1 resistance

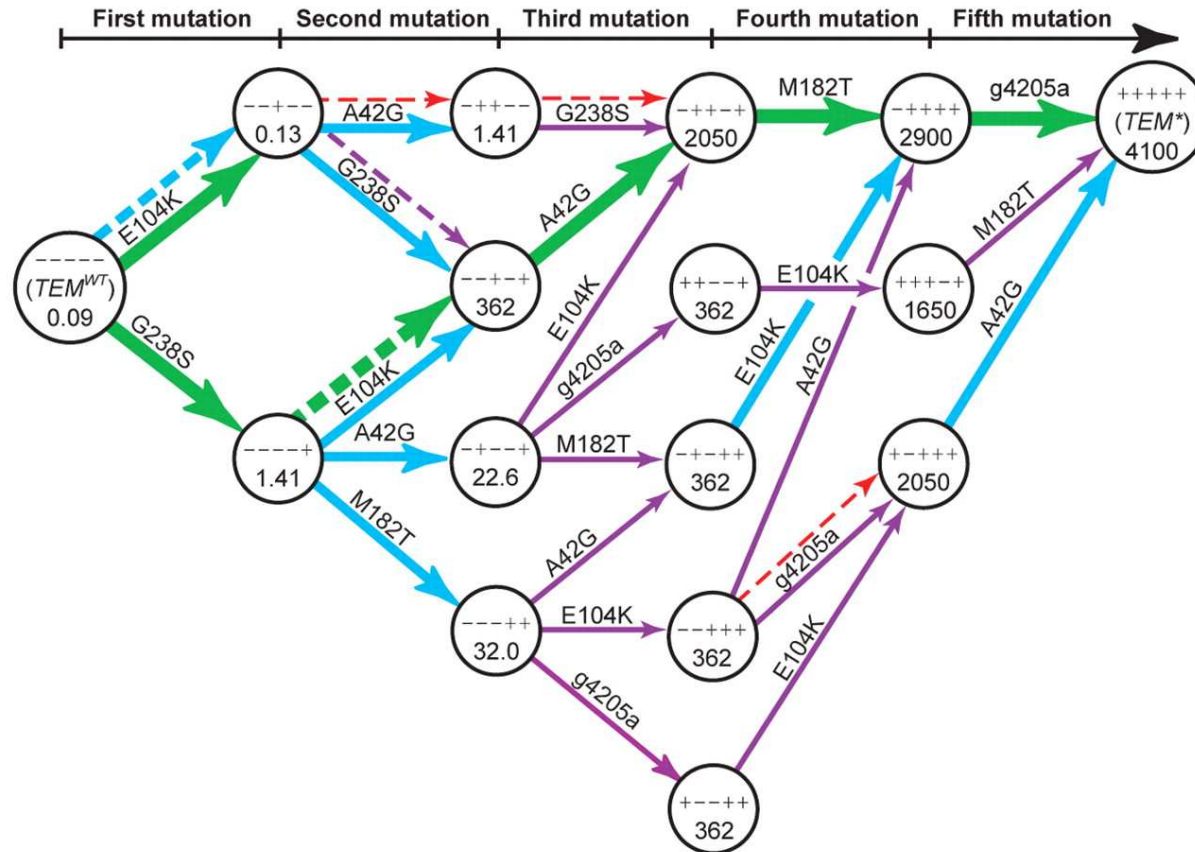
D.M. Weinreich et al., Science **312**, 111 (2006)



- 5 mutations increase the MIC by  $4 \times 10^4$
- Construct all  $2^5 = 32$  combinatorial mutants

# Pathways to TEM-1 resistance

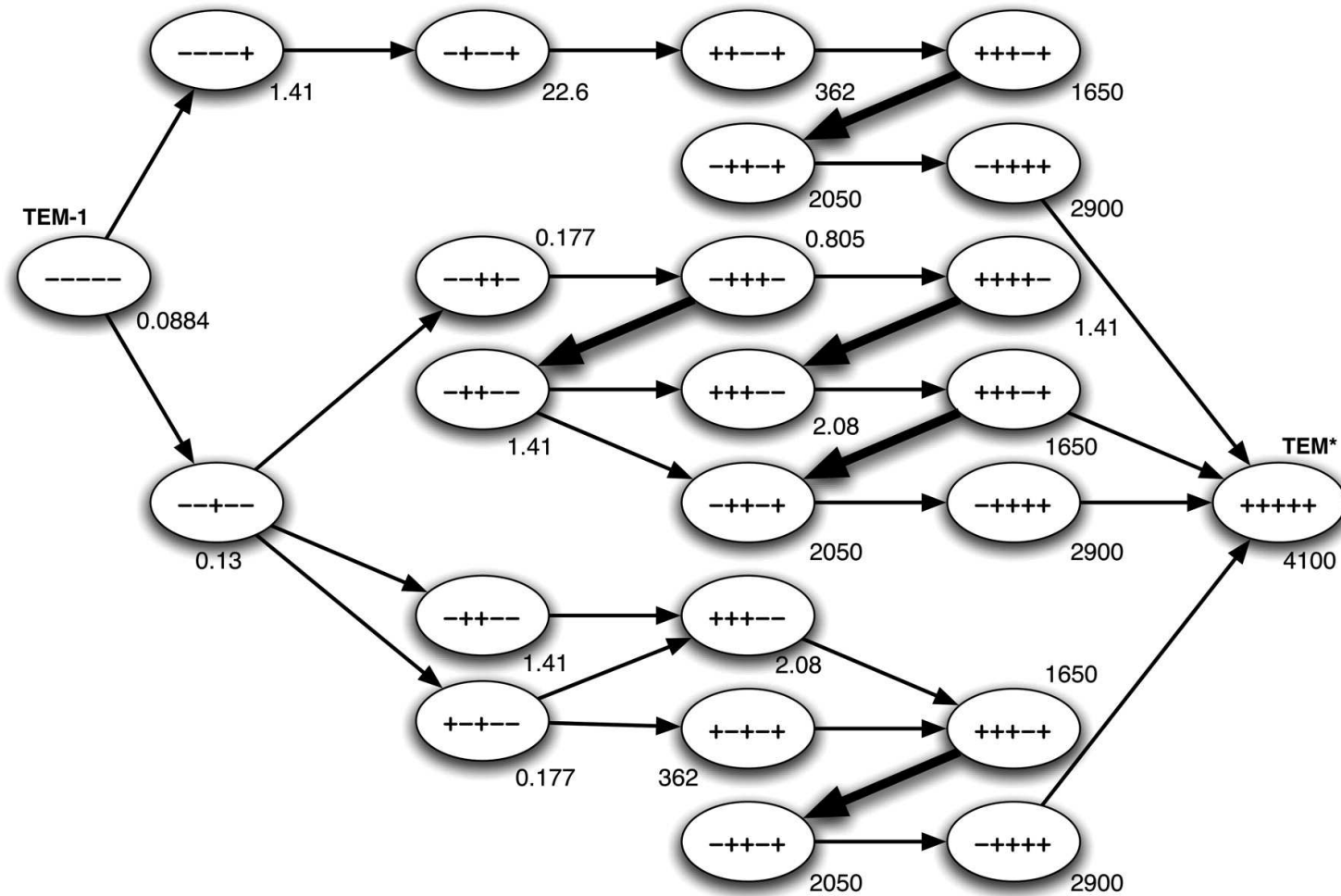
D.M. Weinreich et al., Science **312**, 111 (2006)



- Only 18 out of  $5! = 120$  directed mutational pathways are monotonically increasing in resistance, and only a few of them have appreciable weight

# Pathways to TEM-1 resistance

De Pristo et al. 2007

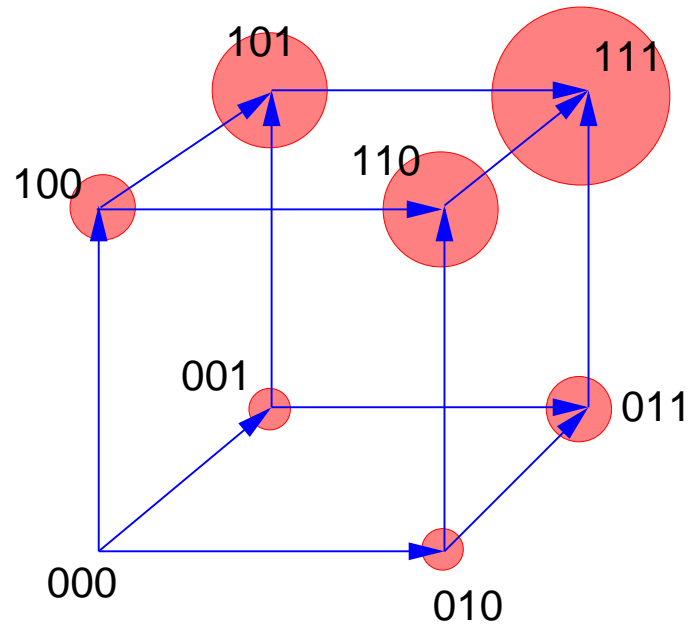


- 27 out of 18651552840 undirected pathways are monotonically increasing



Mathematical framework

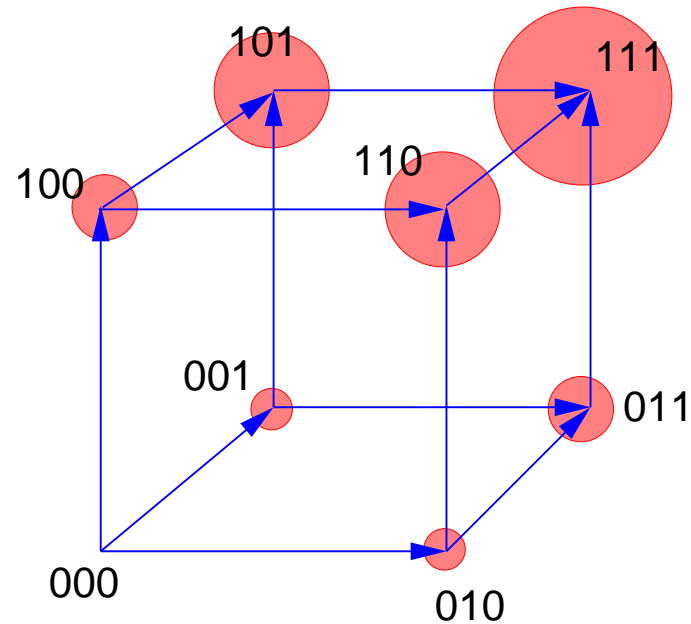
# Pathways in fitness landscapes



- Genotypes are encoded by binary sequences  $(\sigma_1, \dots, \sigma_L)$  where  $\sigma_i = 1$  ( $\sigma_i = 0$ ) denotes the presence (absence) of a mutation at position  $i$
- A **fitness or resistance landscape** is a function on the  $L$ -dimensional hypercube  $\{0, 1\}^L$  of genotypes
- The **fitness graph** is obtained by orienting the links in the direction of increasing fitness

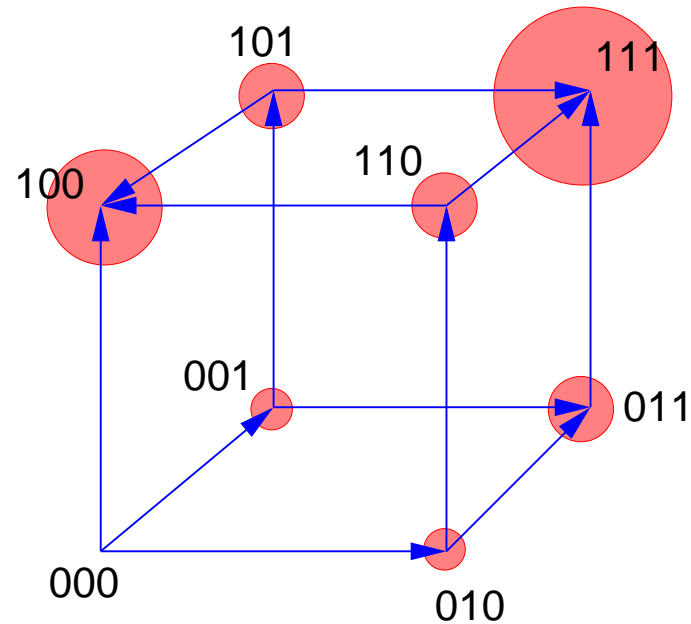
Crona et al. 2013

# Pathways in fitness landscapes



- $L = 3$  mutational steps from the wild type 000 to the adapted mutant 111
- Mutations can occur in  $3 \times 2 \times 1 = 3! = 6$  different orders corresponding to 6 possible directed pathways
- If all mutations are unconditionally beneficial all pathways are **accessible** (= increasing in fitness)

# Pathways in fitness landscapes



- **Sign epistasis** occurs if mutations can be beneficial or deleterious depending on the genetic context Weinreich et al. 2005
- This implies that parallel arrows in the fitness graph point in opposite directions
- Sign epistasis reduces the number of direct accessible paths but may increase the number of evolutionary endpoints

# Accessibility and predictability

- Pathways are **accessible** if fitness/resistance increases monotonically
- Existence of a **small but nonzero** fraction of accessible pathways implies high (retrospective) predictability

# Accessibility and predictability

- Pathways are **accessible** if fitness/resistance increases monotonically
- Existence of a **small but nonzero** fraction of accessible pathways implies high (retrospective) predictability

## Questions for theory

- How does accessibility depend on the structure of the fitness landscape and on the boundary conditions of the paths?
- How typical is it that a small but nonzero fraction of pathways are accessible?

# Accessibility percolation

JK, arXiv:1903.11913

- Null model: assign fitness at random to genotypes Kauffman & Levin 1987
- Probability of existence of accessible paths generically displays a sharp **percolation transition** from 0 to 1 at a critical value  $\beta^*$  of the fitness quantile  $\beta \in [0, 1]$  between initial and final genotype
- For directed paths on the hypercube  $\beta^* = 1 - \frac{\ln L}{L} \rightarrow 1$  for  $L \rightarrow \infty$   
Hegarty & Martinsson 2014
- Mutational reversions increase accessibility such that  $\beta^* < 1$  for  $L \rightarrow \infty$   
Berestycki et al. 2017
- For sequences with  $a$  alleles per site B. Schmiegelt, JK, 2019

$$\beta^* \approx \frac{\ln a}{a} + \frac{1 + \ln a}{a^2} \rightarrow 0 \text{ for } a \gg 1$$

- Near  $\beta = \beta^*$  the number of accessible paths is small and hence predictability is high

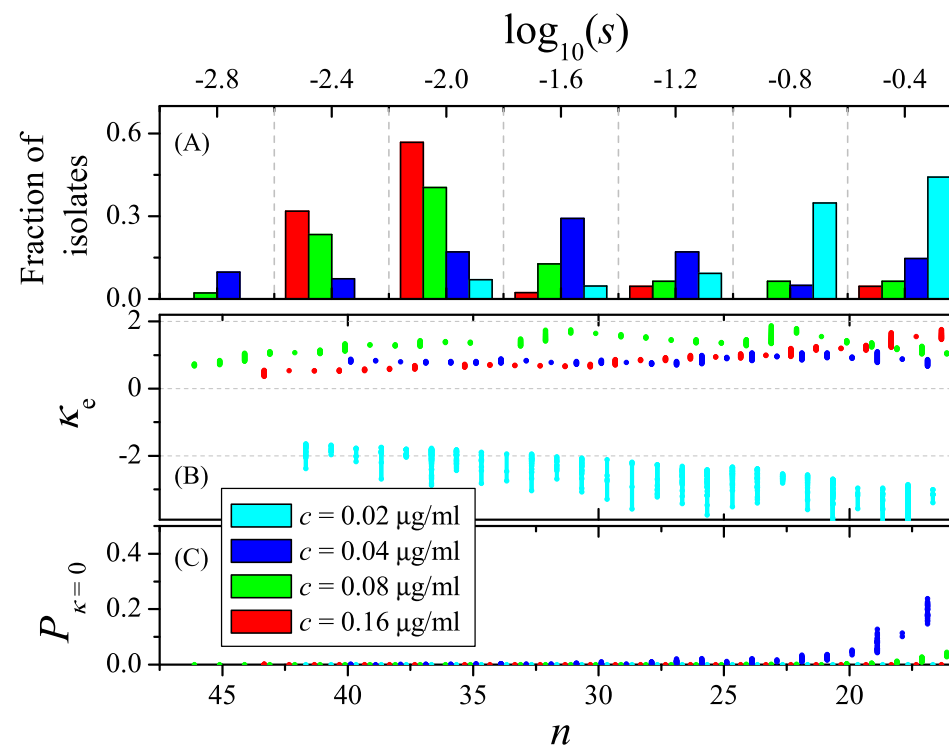
Exploring the TEM-1 resistance landscape



# A panel of resistance mutations

M.F. Schenk et al., PLoS Genet. 2012

- At least **48** out of **2583** point mutations increase resistance against cefotaxime



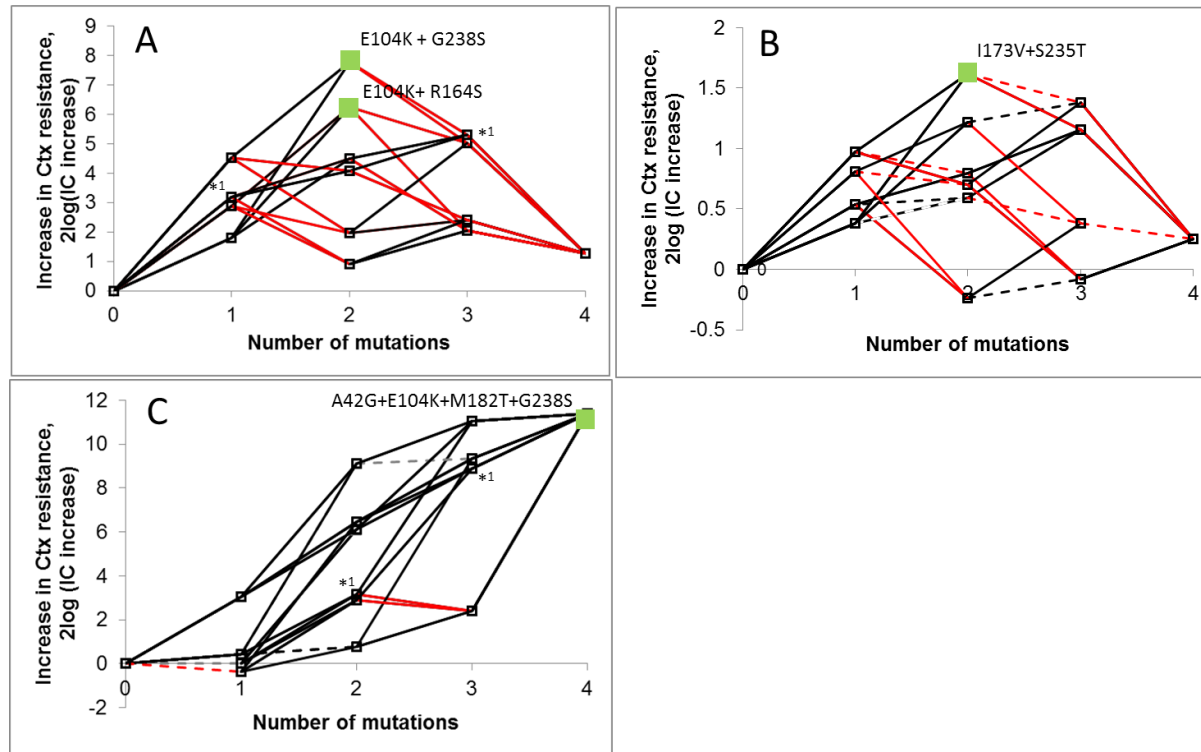
- Analysis using **extreme value theory** yields power law distribution of mutational effects with an exponent  $\sim 1$

# Construction of combinatorial resistance landscapes

- Constructing all possible  $2^{48} \approx 2.8 \times 10^{14}$  combinatorial mutants is obviously unfeasible
- The choice of a **subset** of mutations is expected to bias the structure of the fitness landscape: [de Visser & Krug, Nat. Rev. Genet. 2014](#)
  - singly beneficial vs. singly deleterious mutations
  - mutations chosen for individual or collective effects
  - mutations occurring along an adaptive trajectory
- Here we consider 4-dimensional landscapes constructed from three subsets of **individually beneficial** mutations: [Schenk et al. 2013](#)
  - non-synonymous mutations of strong effect
  - non-synonymous mutations of weak/typical effect
  - synonymous mutations of strong effect

# Mutations chosen for individual vs. collective effect

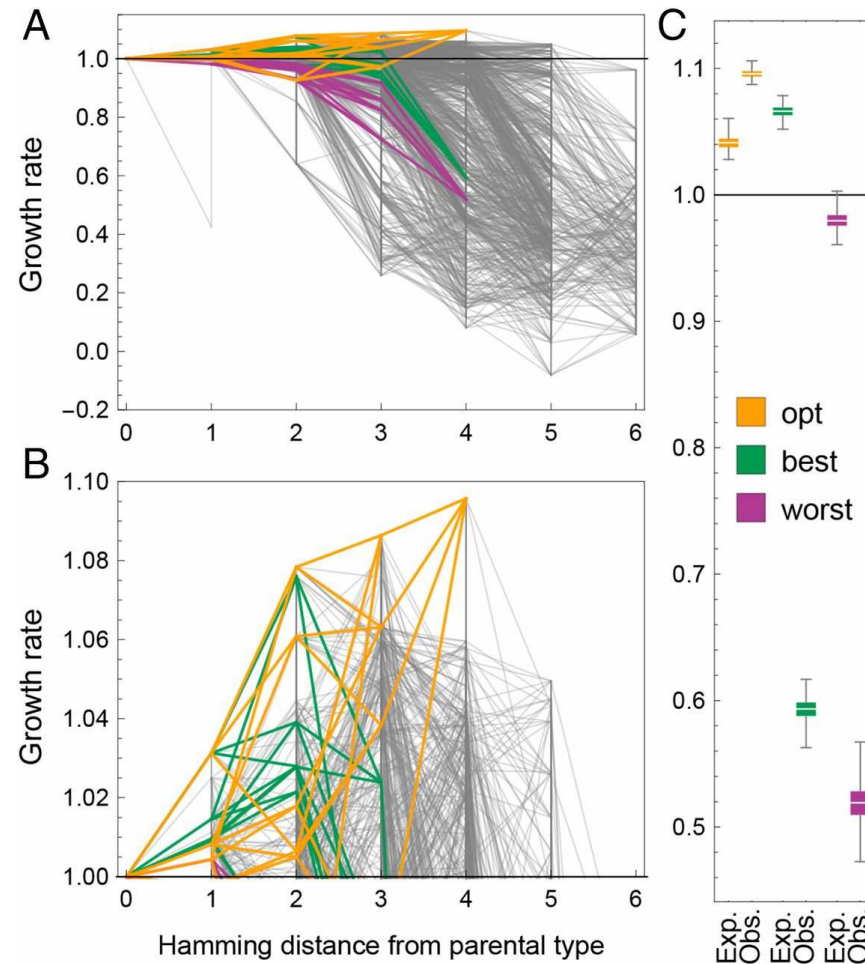
M.F. Schenk et al., Mol. Biol. Evol. (2013)



- Mutations chosen for individual effect interact more strongly and negatively than mutations chosen “with hindsight” because of their collective effect

# Similar patterns observed in yeast

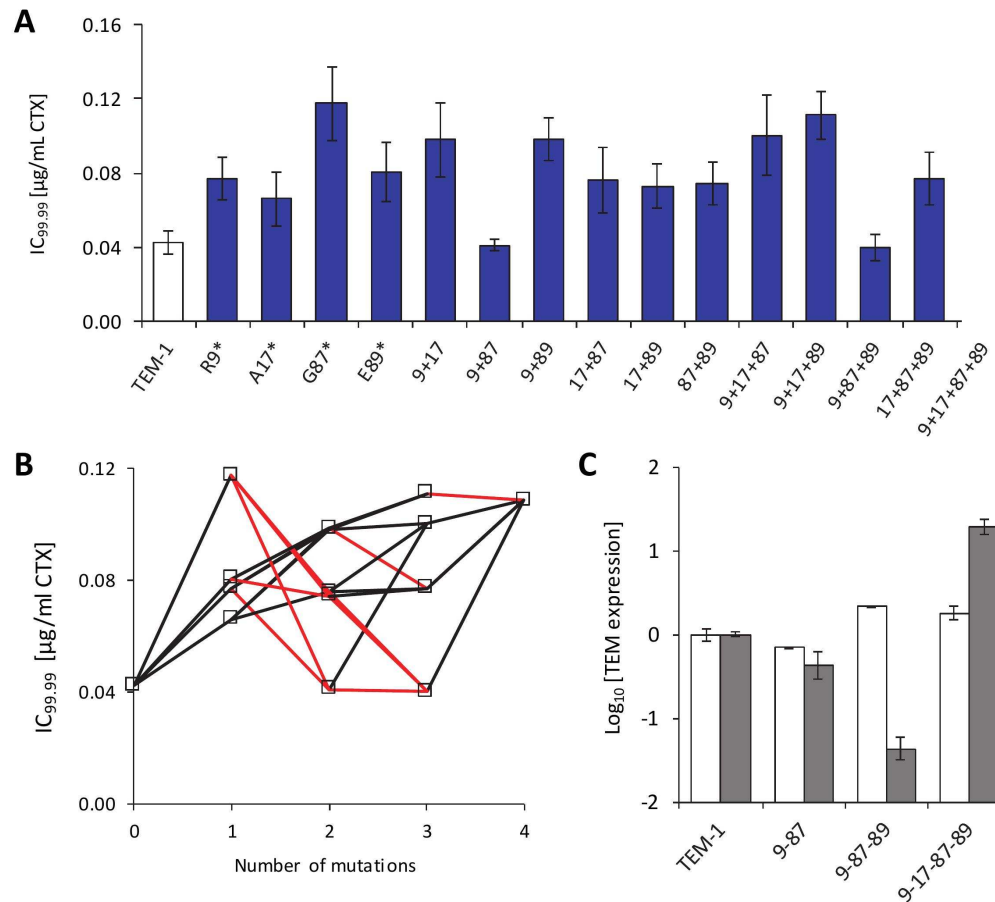
Bank et al., PNAS 2016



- Combinatorial study of 13 mutations in the Hsp90 heat shock protein

# Synonymous resistance landscape

M.P. Zwart et al., Heredity (2018)



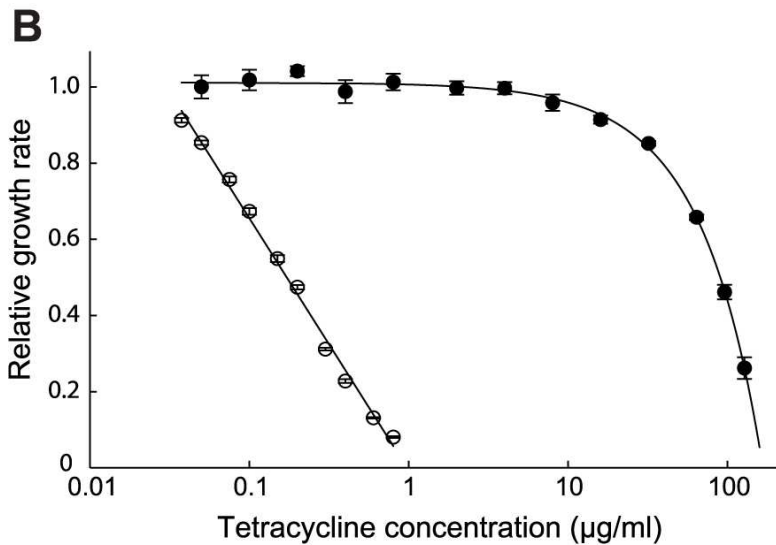
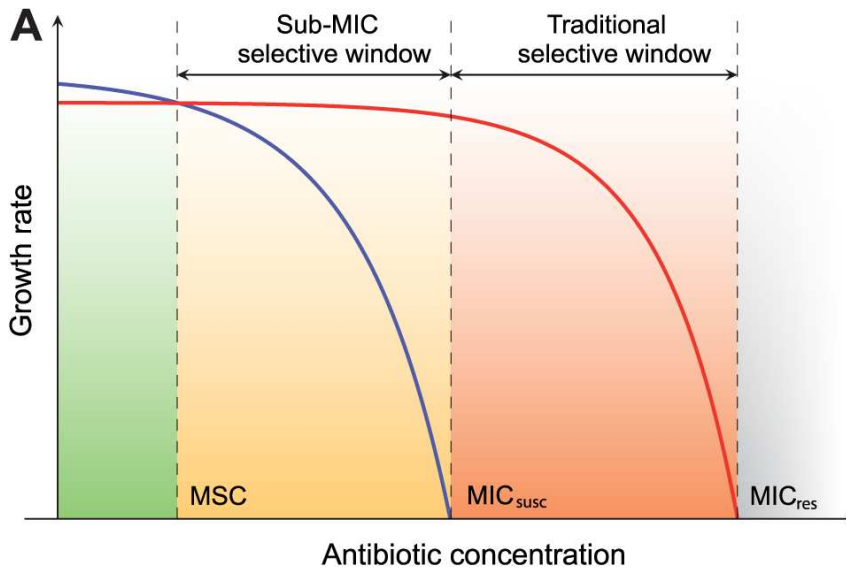
- Landscape displays a “layered” structure that may be related to translational bottlenecks

M. Josupeit, JK, arXiv:2009.10621

# Concentration-dependent fitness landscapes

S. Das, S. Direito, B. Waclaw, R. Allen, JK, eLife 9:e55155 (2020)

# Dose-response curves



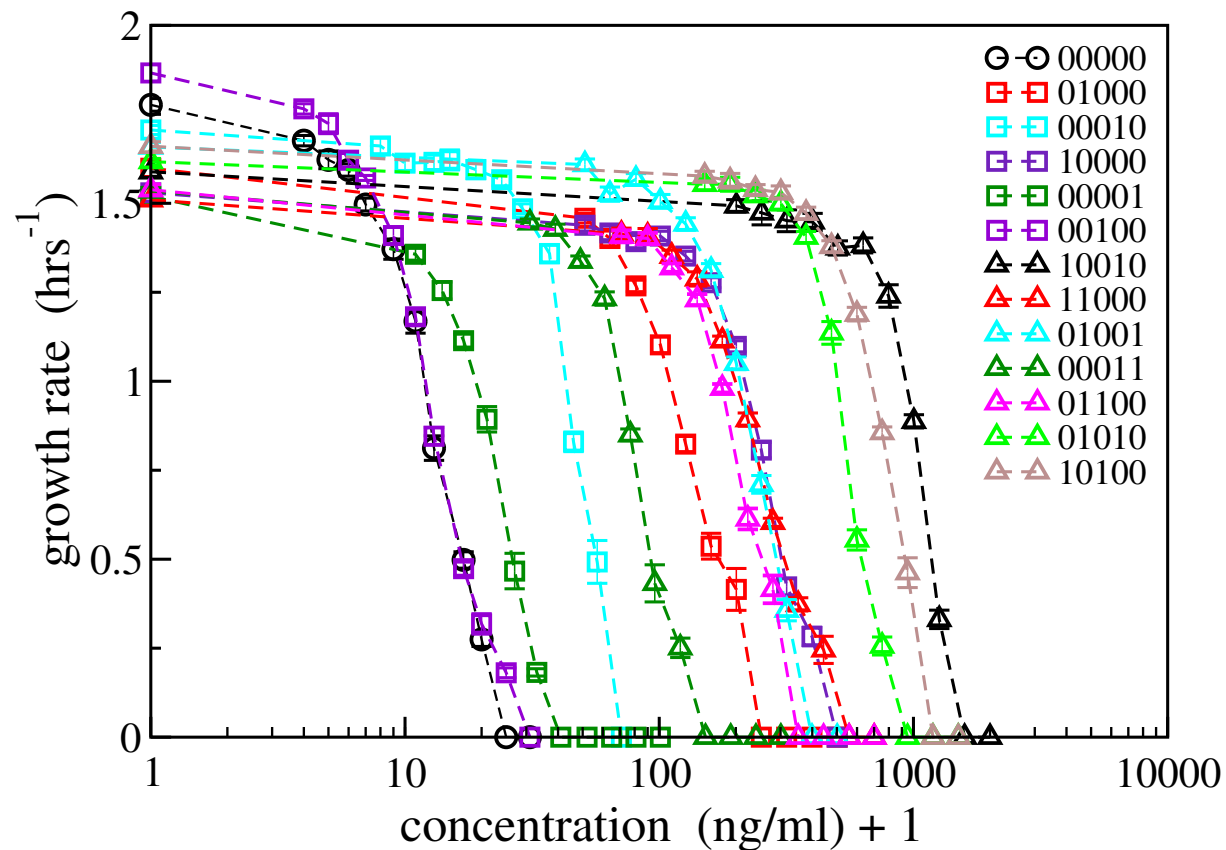
- Mutations that increase resistance often decrease growth rate in the absence of antibiotic (**null fitness**)
- As a consequence the dose-response curves of susceptible and mutant strains cross at the **minimal selective concentration (MSC)**
- The **mutant selection window** is the concentration range

$$MSC < c < MIC_{res}$$

# Observation 1: Scaling of dose-response curves

- Single and double mutations in *Escherichia coli* conferring resistance against ciprofloxacin

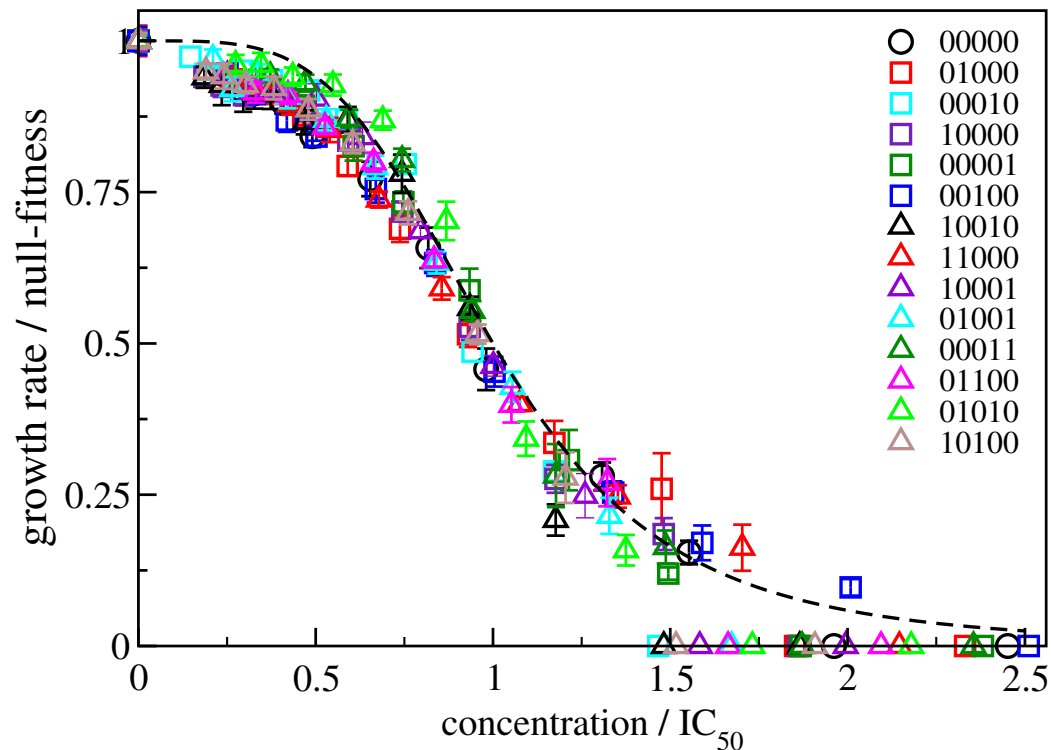
S. Direito, B. Waclaw, R. Allen





# Observation 1: Scaling of dose-response curves

- Single and double mutations in *Escherichia coli* conferring resistance against ciprofloxacin S. Direito, B. Waclaw, R. Allen



- Shape of dose-response curve is a Hill function  $f(x) = (1 + x^4)^{-1}$

## Observation 2: Independent marginal phenotypes

- Null-fitness and MIC of multiple mutants **combine multiplicatively** or display negative interactions data from Marcusson et al., PLoS Pathogens 2009

Strain	String	log(null-fitness)	Non-epistatic	log(MIC)	Non-epistatic
MG1655	00000	0.00 (± .004)		0.00 (± .35)	
LM378	10000	0.01 (± .016)		3.17 (± .70)	
LM534	01000	-0.01 (± .018)		2.75 (± .70)	
LM202	00010	-0.19 (± .020)		0.69 (± .70)	
LM351	00001	-0.094 (± .014)		1.08 (± .70)	
LM625	11000	-0.030 (± .011)	0.0 (± .038)	3.17 (± .70)	5.92 (± 1.1)
LM421	10010	-0.15 (± .019)	-0.18 (± .040)	4.13 (± .70)	3.56 (± 1.1)
LM647	10001	-0.051 (± .013)	-0.084 (± .034)	3.44 (± .70)	4.65 (± 1.1)
LM538	01010	-0.19 (± .020)	-0.20 (± .042)	4.13 (± .70)	3.46 (± 1.1)
LM592	01001	-0.083 (± .015)	-0.10 (± .036)	3.16 (± .70)	3.83 (± 1.1)
LM367	00011	-0.20 (± .026)	-0.28 (± .038)	2.06 (± .70)	1.77 (± 1.1)
LM695	11010	-0.24 (± .017)	-0.19 (± .058)	3.85 (± .70)	6.61 (± 1.1)
LM691	11001	-0.073 (± .013)	-0.094 (± .052)	3.85 (± .70)	7.00 (± 1.4)
LM709	10011	-0.24 (± .027)	-0.274 (± .054)	4.54 (± .70)	4.94 (± 1.4)
LM595	01011	-0.51 (± .051)	-0.294 (± .056)	4.54 (± .70)	4.52 (± 1.4)
LM701	11011	-0.42 (± .037)	-0.284 (± .072)	4.83 (± .70)	7.69 (± 1.8)

- This pattern was first observed for resistance mutations in *Salmonella enterica* and *E. coli* Knopp & Andersson, mBio 2018

# Model with predictable concentration dependence

- $L$  resistance mutations  $i = 1, \dots, L$  characterized by null-fitness  $r_i < 1$  and resistance  $m_i > 1$  relative to the wild type
- Dose-response curve of a mutant  $\sigma = (\sigma_1, \dots, \sigma_L)$  is given by

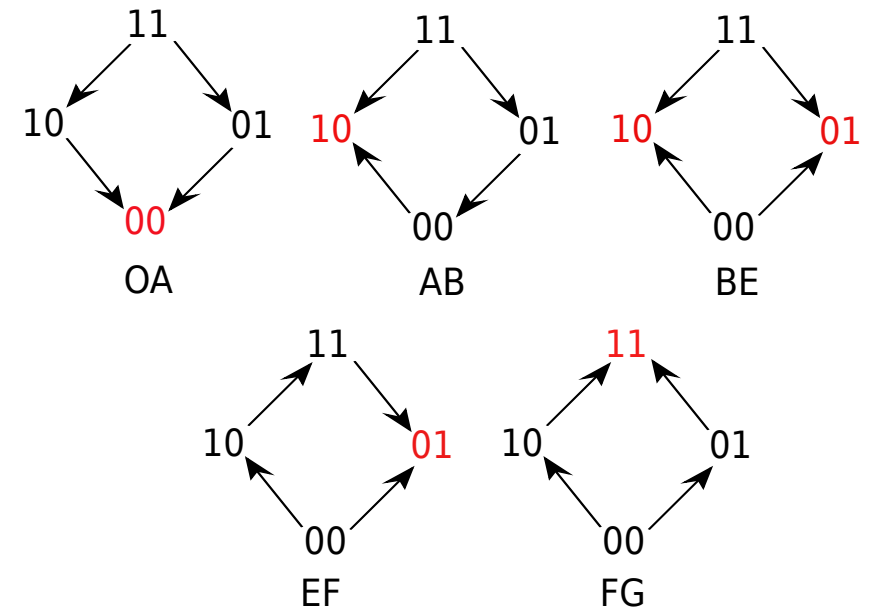
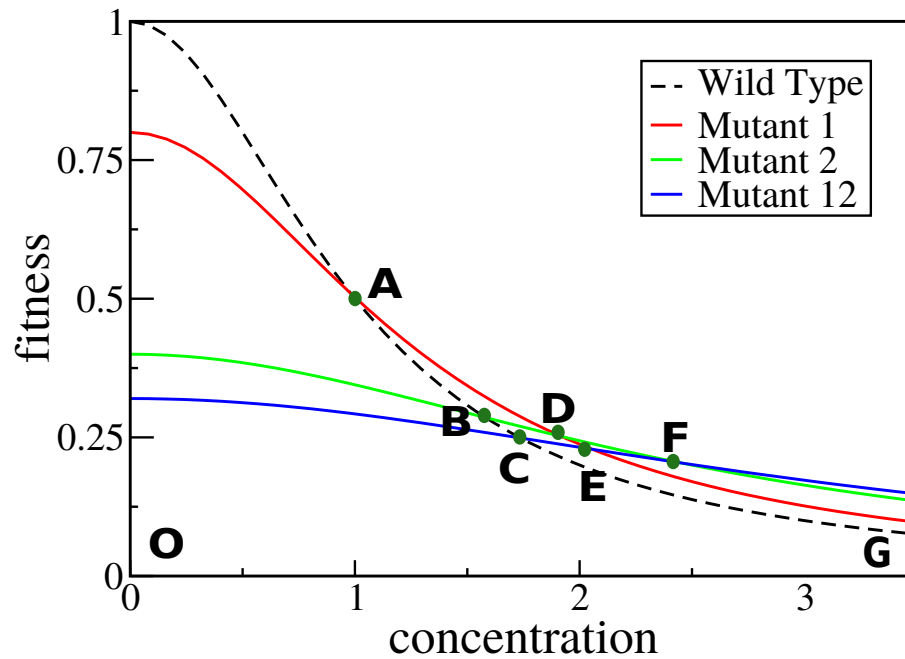
$$w_\sigma(x) = r_\sigma f(x/m_\sigma)$$

where the function  $f(x)$  is independent of  $\sigma$  and the marginal phenotypes combine multiplicatively as

$$r_\sigma = \prod_{j=1}^L (r_j)^{\sigma_j} \quad \text{and} \quad m_\sigma = \prod_{j=1}^L (m_j)^{\sigma_j}$$

- Resistance is quantified by the concentration at which growth drops by 50% ( $IC_{50}$ ), which implies that  $f(1) = \frac{1}{2}$
- In this way  $2^L$  concentration-dependent fitness values can be predicted from  $2L$  single mutant phenotypes and one shape function

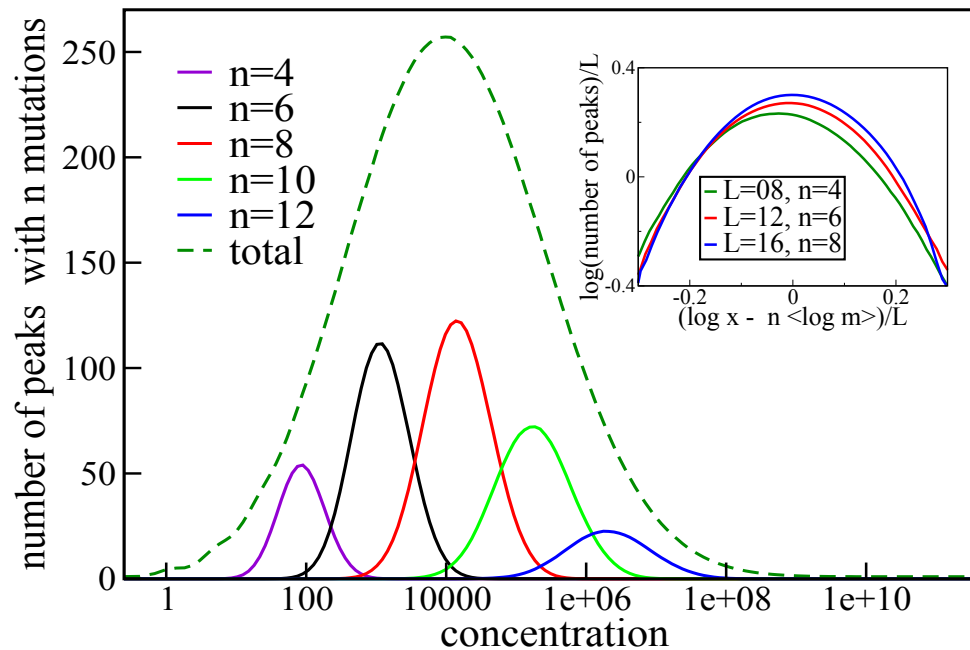
# Two resistance mutations



- Fitness landscape evolves from single-peaked to two-peaked and back
- Not all rank orders can appear in this process

# Maximal ruggedness at intermediate concentrations

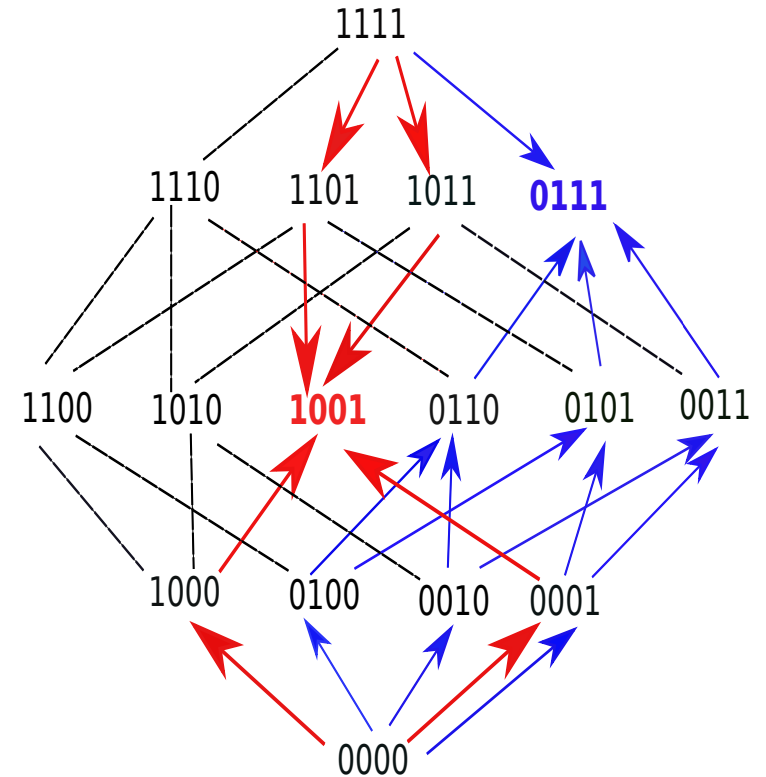
- $L = 16$  mutations with randomly distributed  $r_i, m_i$
- Quantify ruggedness by the number of local fitness peaks



- Typical fitness peaks carry  $n$  mutations at  $\ln x \sim n \langle \ln m_i \rangle$ , and the maximal number of peaks grows exponentially with  $L$

# Landscapes are nevertheless highly accessible

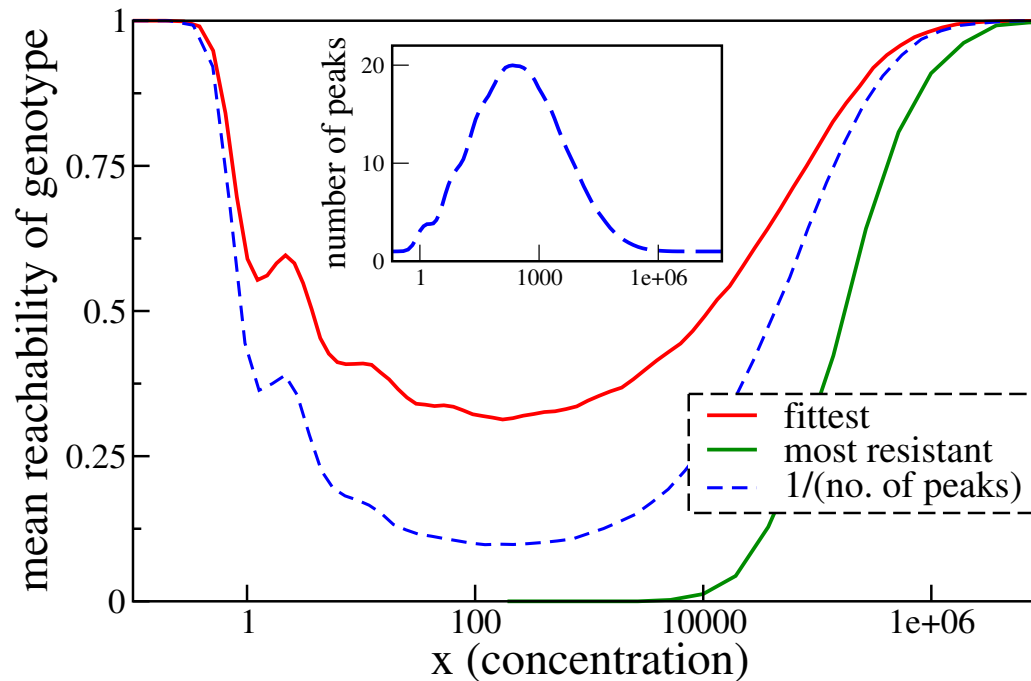
- Scaling and absence of positive marginal epistasis imply that certain rank orders are forbidden at any concentration
- As a consequence, any peak genotype is accessible from all its sub- and supersets
- In particular, the fittest type is **always accessible** from the wild type



Two-peaked landscape at an intermediate concentration

# Reachability of the fittest and most resistant mutant

- $L = 10$  mutations with randomly distributed  $r_i, m_i$



- Probability of reaching the fittest/most resistant mutant from the wild type using strong selection/weak mutation dynamics

# Summary

- Evolution of antimicrobial resistance through multiple mutational steps is a model for evolutionary predictability
- Combinatorial construction of resistance landscapes reveals a systematic dependence on the choice of the combined mutations
- Tradeoff between resistance and growth rate induces rugged fitness landscapes at intermediate antibiotic concentration
- Despite their ruggedness these landscapes are remarkably accessible and the evolution of high levels of resistance remains facile
- **Outlook:** Consider time-dependent antibiotic concentrations as a model for evolution in changing environments



# Thanks to

- **Cologne:**

Jasper Franke, Stefan Nowak, Benjamin Schmiegelt, Ivan Szendro, Sungmin Hwang, Suman Das

- **Wageningen:**

Martijn Schenk, Mark Zwart, Manja Saebelfeld, Arjan de Visser

- **Edinburgh:**

Susana Direito, Bartek Waclaw, Rosalind Allen